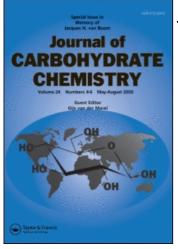
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Synthesis of Sugars From D-Ribonolactone. II. An Alternative Synthesis of D-Erythrose

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SYNTHESIS OF SUGARS FROM $\stackrel{\text{D-RIBONOLACTONE.}}{=}$ II.

AN ALTERNATIVE SYNTHESIS OF <u>D</u>-ERYTHROSE

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ABSTRACT

<u>D</u>-Erythrose was synthesized in four steps from <u>D</u>-ribono-1,4-lactone via the 3,5-<u>O</u>-benzylidene derivative of the latter compound. Reduction of the benzylidene <u>D</u>-ribonolactone, and periodate cleavage of the resulting 3,5-<u>O</u>-benzylidene-<u>D</u>-ribitol were performed in a one-flask reaction. The ensuing 2,4-<u>O</u>-benzylidene-<u>D</u>-erythrose was hydrolyzed with 10% acetic acid to obtain syrupy <u>D</u>-erythrose.

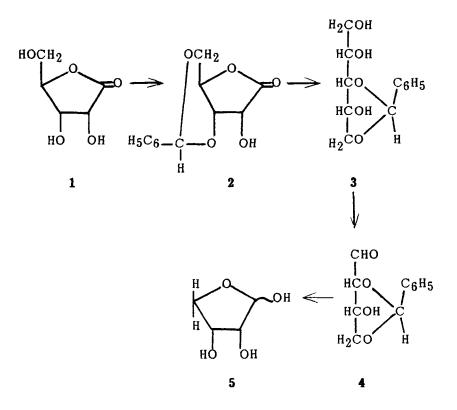
INTRODUCTION

In continuation of our efforts to utilize <u>D</u>-ribono-1,4-lactone (1) as starting material for the synthesis of 4-, 5-, and 6-carbon normal and deoxy sugars, we describe here the synthesis of <u>D</u>-erythrose (5). We have reported¹ the synthesis of <u>L</u>-erythrose from 1 via its 2,3-<u>O</u>-isopropylidene derivative. A number of methods are available in the literature for the preparation of 5. These include i) Ruff degradation^{2,3} of calcium <u>D</u>arabinonate; ii) ozonolysis³ of 1,4,6-tri-<u>O</u>-acetyl-2,3-dideoxy-<u>D</u>-erythrohex-2-enopyranose, followed by hydrolysis of the acetyl groups with aqueous hydrochloric acid; iii) oxidative cleavage³ of methyl β -<u>D</u>glucofuranoside 5,6-carbonate with lead tetraacetate, and removal of the carbonate group with aqueous barium hydroxide; iv) periodate cleavage of a) 4,6-O-benzylidene-D-glucose⁴ or -D-glucitol,⁴ b) 1,3-O-ethylidene-D-mannitol,⁵ or c) 4,6-O-ethylidene-D-glucose⁶⁻⁸ or -D-glucitol,⁹ followed by hydrolysis with aqueous sulfuric acid of the resultant 2,4-O-benzylidene-(4) or -ethylidene-D-erythrose; v) MacDonald-Fischer^{10,11} degradation of D-arabinose diethyl dithioacetal;¹² vi) oxidative cleavage of D-glucose with two equivalents of lead tetraacetate, followed by hydrolysis of the 3,4-di-O-formyl-D-erythrose with aqueous hydrochloric acid;¹³ and vii) borohydride reduction followed by periodate cleavage of 3,4-Oisopropylidene-D-arabinose to obtain the 2,3-O-isopropylidene-Derythrose.¹⁴

Of these, the method preferred most frequently for the preparation of 5 is the one involving the periodate cleavage of $4,6-\underline{O}$ -ethylidene- \underline{D} glucose.⁶⁻⁸ In contrast, the method⁴ employing the analogous $4,6-\underline{O}$ benzylidene- \underline{D} -glucose has been largely ignored for this purpose. The reason for this is that, whereas the latter compound can be prepared¹⁵ at best in 42% yield after considerable effort, yields of 70-87% can be achieved^{8,16,17} more readily in the preparation of the former derivative. Recently, a high-yield synthesis of 3,5- \underline{O} -benzylidene- \underline{D} -ribono-1,4lactone¹⁸ (2) has been described,¹⁹ which makes the preparation of \underline{D} erythrose (5) via the 2,4- \underline{O} -benzylidene derivative **4** more attractive.

DISCUSSION

The route to 4 involved the same sequence of reactions that we and others have followed for the synthesis of 2,3-Q-isopropylidene- $\underline{L}^{-1,20,21}$ and $-\underline{D}$ -erythrofuranose.¹⁴ Accordingly, 2, prepared by the literature procedure,¹⁹ was converted to 4 in a one-flask reaction by reduction with sodium borohydride in aqueous ethanol followed by oxidative cleavage of the intermediate 3,5-Q-benzylidene- \underline{D} -ribitol (3) with periodate; the yield of 4 was 63% overall. The intermediate 3 was isolated from a different, small-scale run and shown to be identical with 3 prepared by reduction of 2 with lithium aluminum hydride, as judged by thin-layer chromatography (TLC).²² The 2,4-Q-benzylidene- \underline{D} -erythrose (4) was essentially homogeneous according to TLC, and its mobility was



quite different from that of both 2 and 3. The infrared (IR) spectrum of 4 showed all the requisite peaks except for the carbonyl signal for the aldehyde group, which was very weak. However, 4 showing strong carbonyl absorption at 1715 cm⁻¹, and all the other expected peaks, was obtained from another, identical run in which the work-up procedure was slightly modified (see experimental section). Chromatographically, the products (4) from the two runs were identical; their IR spectra differed only in the relative intensities of some of the peaks. It (4) showed a mutarotation of small magnitude that is reminiscent of that reported by Schaffer⁷ for 2,4-Q-ethylidene-<u>D</u>-erythrose in water $[[\alpha]_D -40^\circ -> -43.5^\circ]$. It is not clear whether equilibria between the monomeric and oligomeric forms of 4 or different anomers of the oligomeric forms, or both are responsible for its mutarotation.

Baggett et al.²³ have suggested a dimeric structure for 4^4 on the basis that it showed neither a carbonyl absorption in the IR nor an

aldehydic proton signal in the ¹H NMR, as well as by analogy with the proposed⁷ dimeric structure for 2,4-<u>O</u>-ethylidene-<u>D</u>-erythrose. However, the monomeric structure for 4⁴ was preferred by Thiem and Wessel²⁴ on the basis of methylation studies. Our observation of the TLC behavior of 4 suggests that it migrates as a monomer; its mobility relative to 2 and 3 was 1.5 and 2.1, respectively. For it to possess the dimeric structure, its R₂ and R₃ values would have been expected to be much smaller than observed. The fact that strong carbonyl absorption was observed by us for a preparation of 4 is highly supportive of its monomeric structure. The monomeric structure would also seem to be supported by the fact noted by Baggett et al.²³ that 4 reduces Fehling's solution whereas its ethylidene counterpart does not.

Sowden⁴ removed the benzylidene group of 4 by hydrolysis with 0.1 N sulfuric acid at reflux temperature. We preferred to employ 10% acetic acid for this purpose in order to obviate the need for extra measures for removal of the sulfate ions. The hydrolysis of 2,4-Q-benzylidene-D-threose under these conditions has been reported by Neish.⁹ Accordingly, 4 was hydrolyzed with 10% acetic acid, and 5 was obtained as a colorless syrup in 83% yield. This product was identical with authentic D-erythrose (purchased commercially) and the L-erythrose (also prepared¹ from 1) according to TLC. It mutarotated in water from a low to high value, reaching equilibrium after six days with an [α]_D of -38.4°. The direction of mutarotation observed here is in agreement with that reported^{2,3,12} for 5 in the literature. Previously we have reported¹ an [α]_D of +36.8° (water) for the L-erythrose, which was also prepared from

1. These two values are within the $\pm 3^{\circ}$ of 38° that Baxter and Perlin²⁰ suggested as the correct optical rotation of erythrose [(+) for the <u>L</u> and (-) for the <u>D</u>]. This suggestion²⁰ is corroborated by the fact that we have prepared the two enantiomers of erythrose from the same starting material (1) whose specific rotations fall within this range and which are opposite in sign.

Perlin and Brice¹³ postulated the formation of oligomers of 5 at low temperature to explain the reported, low specific rotations for 5 and its enantiomer, and to account for the presence of more than one component on paper chromatograms. These investigators showed¹³ that keeping a solution of 5 frozen for 18 hours and then thawing lowers the specific rotation from -31° to -6°, the original value being regained after four hours at room temperature; this rotatory change was accompanied by an increase in the slower moving component at the lower temperature. Andersson, Theander, and Westerlund²⁵ noted the presence of signals from the anomeric protons of monomeric and oligomeric 5 in the 100-MHz ¹H NMR spectrum in deuterated acetate buffer, pH 4.5. In keeping with the postulate¹³ of Perlin and Brice, these investigators determined²⁵ from ¹H NMR studies that heating of the solution increases the proportion of monomeric 5. The presence of oligomers of 5 in concentrated solutions or in solutions freshly prepared from syrups was also observed by Serianni, Clark, and Barker²⁶ from ¹³C NMR studies. The optical rotatory and chromatographic behavior of 5 synthesized here are consistent with these findings.^{13,25,26}

Thus, the method described here should provide an excellent alternative to the existing procedures for the preparation of \underline{D} -erythrose (5) of high purity in good yield from a readily available starting material (1).

EXPERIMENTAL

Melting points were determined with a Fisher-Johns apparatus and are uncorrected. Optical rotations were measured at ambient temperature in a 1-dm cell with a Perkin-Elmer Model 241 automatic polarimeter. IR spectra were recorded with a Perkin-Elmer Model 283B infrared spectrophotometer. TLC was performed with silica gel G in 10:1 chloroform-methanol (solvent <u>A</u>) or 3:1:1 ethyl acetate-acetic acid-water (solvent <u>B</u>). Evaporations were performed under reduced pressure at 40-45 °C. Benzaldehyde was used as received from the supplier. Authentic D-erythrose was purchased from Sigma Chemical Co., St. Louis, Missouri.

<u>3,5-O-Benzylidene-D-ribono-1,4-lactone</u> (2). This was prepared in 91.5% yield essentially as described¹⁹ by Chen and Joullie, with the modification that the crude product was washed with <u>n</u>-pentane prior to washing with 5% sodium hydrogen carbonate, water, and n-pentane; m.p.</u> 230-232 °C [lit.^{18,19} m.p. 233-235.5 °C and 230-231.5 °C, respectively]; $[a]_D$ -173.2° (<u>c</u> 0.45, <u>N,N</u>-dimethylformamide) [lit.^{18,19} [$a]_D$ -174.1° and -177.0°, respectively (both in <u>N,N</u>-dimethylformamide)]; R_F 0.42 (solvent A). The crude product was used directly without recrystallization.

<u>2,4-O-Benzylidene-D-erythrose</u> (4). To a suspension of 2 (2.36 g, 10 mmol) in a mixture of ethanol (120 mL) and water (40 mL) was added over 2 min a solution of sodium borohydride (1.4 g, 37.8 mmol) in water (50 mL). The insoluble solid dissolved over the next 45 min to give a clear solution [the mixture gradually becomes turbid again, presumably due to the separation of the intermediate 3,5-O-benzylidene-D-ribitol (3)]. After 24 h, the pH was adjusted to approximately 6 with 20% aqueous acetic acid, which resulted in the dissolution of the separated material.

A solution of sodium metaperiodate (2.25 g, 10.5 mmol) in water (20 mL) was added over 5 min during which time a solid (inorganic salts) separated as fine needles. After 2 h at room temperature, the reaction mixture was evaporated, the solid gradually dissolving as the ethanol evaporated. The colorless, crystalline residue was dried by evaporation with ethanol (2 x 50 mL) and then was extracted with hot ethyl acetate (350 mL in several portions). The combined extracts were washed with water (2 x 25 mL), dried (sodium sulfate), and evaporated to obtain light yellow, syrupy 4 (1.31 g, 63%), which was essentially homogeneous according to TLC; [α]_D -65.3° (3 min) \rightarrow -62.6° (equilibrium, 3 h) (<u>c</u> 0.65, chloroform), -68.5° (<u>c</u> 0.27, <u>N,N</u>-dimethylformamide) [lit.²³ [α]_D -20° (<u>N,N</u>-dimethylformamide)]; R_F 0.64 (major), 0.02 and 0.0 (traces) (solvent <u>A</u>); IR (film): 3400 (OH), 1720 (vw, aldehyde carbonyl), 1105, 1075, 1030 (sh), and 1010 (C-O, C-O-C), 768 and 708 cm⁻¹ (monosubstituted phenyl).

In another identical run, the inorganic solid which precipitated during the periodate reaction was filtered off and washed with 1:1 ethanol-water and finally with ethanol. The combined filtrate and washings were evaporated to dryness, and the residue was extracted with hot ethyl acetate; the filtered inorganic solid was also extracted with hot ethyl acetate. All the extracts were combined and dried (sodium sulfate), washing with water being omitted. Concentration of the extracts gave 4 as a pale yellow syrup (86% yield) which was identical with the above product on TLC. In the IR this product showed a strong aldehyde carbonyl peak at 1715 cm⁻¹; otherwise its IR spectrum was identical to that of the above product, differing from it only in the relative intensities of some of the peaks.

<u>D-Erythrose</u> (5). A solution of 4 (1.23 g) in 10% aqueous acetic acid (30 mL) was heated at 100 °C for 1 h. The pale yellow solution was evaporated to dryness. The residual syrup was dissolved in water (15 mL), and the solution was washed with ether (3 x 10 mL) to remove traces of benzaldehyde. The aqueous layer was decolorized with Norit A and evaporated to obtain 5 as a colorless, thick syrup (0.59 g, 83%); $[\alpha]_D$ -15.8° (5 min) -> -36.8° (24 h) -> -38.4° (equilibrium, 6 days) (<u>c</u> 0.77, water) [lit.2,3,5,7,12,13 $[\alpha]_D$ -14.5°, -18.5°, -17.0°, -41.0°, -23.1°, and -32.7°, respectively]; R_F 0.50 (major, monomer), 0.20 (minor, oligomer) (Solvent <u>B</u>); authentic <u>D</u>-erythrose and <u>L</u>-erythrose¹ showed similar spots.

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